





Patient's Identification Code:
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Introduction

This standardized Case Report Form (CRF) is the result of an ongoing effort between the World Health Organization (WHO), The Pan-American Health Organization (PAHO), Institute Pasteur (IP), and the networks of ISARIC, CONSISE PREPARE and REACTing to generate standardized clinical and epidemiological research tools.

DESIGN OF THIS CASE REPORT FORM (CRF)

There are sets of Case Report Forms (CRFs) to be used in combination for prospective cohort studies or case control studies. These sets of CRFs are to be used at admission and at discharge/going home. For any patients admitted for more than 24 hours, the Baseline and Outcome CRF and the Laboratory Results CRF can be copied and used for daily data recording.

For all studies, we recommend completing a minimum of the Child Baseline and Outcome (CBO) CRF, follow by Child Acute Symptoms (CAS). If the patient is admitted to a hospital or has further investigations, complete Child Hospital Stay (CHS) and Child Laboratory Results (CLR) CRFs. We recommend completing the Neonatal CRF and the Maternal Baseline and Outcome CRF to capture maternal and/or neonatal risk factors. If the patient is admitted to an Intensive Care Unit or Pediatric Intensive Care Unit, complete Child Intensive Care (CIC) as well. For follow up visit(s) complete Child follow up visit(s) (CFU).

Complete the outcomes sections in the CBO CRF once all diagnostics laboratory results and final diagnosis are available.

HOW TO USE THIS CRF

When completing the CRF modules, please make sure that:

- The patient or consultee/guardian/representative has been given information about the study and the informed consent form has been completed and signed.
- The study ID codes have been assigned for the patient as per hospital protocol and guidelines.
- The study ID codes should be filled in on all pages of paper CRF forms, all information should be kept confidential at all times, and no identifiable information is recorded on the CRFs.
- Patient's hospital ID and contact details are recorded on a separate contact list to allow later follow up. The contact forms must be kept separate from the CRFs at all times and kept in a secure location.

Each site may choose which data to collect based on available resources and the number of patients enrolled to date. Ideally, data on patients will be collected using all CRF modules as appropriate.

Sites with very low resources or very high patient numbers may select **Child Baseline and Outcome (CBO)** CRF module only. The decision is up to the site Investigators and may be changed throughout the data collection period. All high quality data are valuable for analysis.

GENERAL GUIDANCE

- The CRFs are designed to collect data obtained through patient examination, for patient or parent/guardian/representative interview and review of hospital notes.
- Patient ID codes should be filled in on all pages of paper CRF forms.
- Complete every line of every section, except for where the instructions say to skip a section based on certain responses.
- Selections with square boxes (\square) are single selection answers (choose one answer only). Selections with circles (0) are multiple selection answers (choose as many answers as are applicable).
- It is important to indicate when the answer to a particular question is not known. Please mark the 'Unknown' box if this is the case.
- Some sections have open areas where you can write additional information. To permit standardized data entry, please avoid writing additional information outside of these areas.
- We recommend writing clearly in black or blue ink, using BLOCK-CAPITAL LETTERS.
- Place an (X) when you choose the corresponding answer. To make corrections, strike through (----) the data you wish to delete and write the correct data above it. Please initial and date all corrections.
- Please contact us, if we can help with any CRF completion questions, if you have comments and to let us know that you are using the forms. Please contact Dr Gail Carson by email: gail.carson@ndm.ox.ac.uk







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Disclaimer: These CRFs are intended for use as a standardized document for the collection of clinical data in studies investigating the Zika virus. Responsibility for use of these CRFs rests with the study investigators. ISARIC and the authors of the CRF accept no responsibility for the use of the CRF in an amended format nor for the use of the standardized CRF outside its intended purpose. Formatting issues are in the process of being resolved. Word documents are available in order to adapt and translate the CRFs, however, there may be issues between Macs and PCs. The PDF format is also available, which should be well formatted on both systems.

1) LABORATORY RESULTS

Record all values available ≤24 hours of presentation/admission as part of the clinical care of patient included in the study. Please use standard (SI) units if possible. Please specify the unit used for each result. For repeat testing, use the most abnormal value per day copy page and ensure date of testing and patient IDs are indicated on each page.

	-		,			
1. Date of sampling	/ / 20					
(dd/mm/yyyy):		Value				
Test			Specify unit,	Specify unit, if other specify unit use		
Inflammatory markers						
2. C-reactive protein	☐Yes ☐Not done ☐Unknow	n	□mg/L		□other:	
3. Erythrocyte	☐Yes ☐Not done ☐Unknow	n	□mm/hr		□other:	
sedimentation rate						
4. Procalcitonin	☐Yes ☐Not done ☐Unknow	n	□ng/mL		□other:	
Hematology						
5. Hemoglobin	☐Yes ☐Not done ☐Unknow	n	□g/L	□g/dL	□other:	
6. Hematocrit	☐Yes ☐Not done ☐Unknow	n	□%		□other:	
7. RBC count	☐Yes ☐Not done ☐Unknow	n	□x10 ⁹ /L	□x10³/μL	□other:	
8. MCV	☐Yes ☐Not done ☐Unknow	n	□μm³		□other:	
9. White blood cell	☐Yes ☐Not done ☐Unknow	n	□x10 ⁹ /L	□x10³/μL	□other:	
count						
10. Neutrophils	☐Yes ☐Not done ☐Unknow	n	□10³/mm³	□%	□other:	
11. Lymphocytes	☐Yes ☐Not done ☐Unknow	n	□10³/mm³	□%	□other:	
12. Monocytes	☐Yes ☐Not done ☐Unknow	n	□10³/mm³	□%	□other:	
13. Eosinophils	☐Yes ☐Not done ☐Unknow	n	□10³/mm³	□%	□other:	
14. Basophils	☐Yes ☐Not done ☐Unknow	n	□10³/mm³	□%	□other:	
15. Platelets	☐Yes ☐Not done ☐Unknow	n	□x10 ⁹ /L	□x10³/μL	□other:	
16. APTT	☐Yes ☐Not done ☐Unknow	n	□seconds			
17. PT (seconds)	□Yes □Not done □Unknow	n	□seconds			
18. Blood film	□Yes □Not done □Unknow	n	Describe resi	ults:		
Biochemistry						
19. Urea nitrogen	☐Yes ☐Not done ☐Unknow		□mmol/L	□mg/dL	□other:	
20. Creatinine	☐Yes ☐Not done ☐Unknow	n	□μmol/L	□mg/dL	□other:	
21. Sodium	☐Yes ☐Not done ☐Unknow	n	□mmol/L		□other:	
22. Potassium	☐Yes ☐Not done ☐Unknow	n	□mmol/L		□other:	
23. Total protein	□Yes □Not done □Unknow	n	□g/dL		□other:	
24. Albumin	☐Yes ☐Not done ☐Unknow	n	□g/L		□other:	
25. Bilirubin	□Yes □Not done □Unknow	n	□μmol/L	□mg/dL	□other:	
26. AST/SGOT	☐Yes ☐Not done ☐Unknow	n	□U/L		□other:	
27. ALT/SGPT	□Yes □Not done □Unknow	n	□U/L		□other:	







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28. GGT	□Yes □Not done □Ur	nknown	□U/L	□other:
29. ALP	□Yes □Not done □Ur	nknown	□U/L	□other:
30. Calcium	☐Yes ☐Not done ☐Ur	nknown	□mmol/L	□other:
31. Phosphate	☐Yes ☐Not done ☐Ur	nknown	□mg/dL	□other:
32. Magnesium	☐Yes ☐Not done ☐Ur	nknown	□mmol/L	□other:
33. Amylase	☐Yes ☐Not done ☐Ur	nknown	□U/L	□other:
34. Glucose	☐Yes ☐Not done ☐Ur	nknown	□mmol/L □mg/d	L 🗆 other:
35. Creatine kinase	□Yes □Not done □Ur	nknown	□U/L	□other:
36. Other biochemistr	r y □Yes □Not done □Ur	nknown		
result (specify):			□Unit:	
Other biochemistry	☐Yes ☐Not done ☐Ur	nknown		
result (specify):			□Unit:	
•	available as part of routine ca	•		
If yes, complete table 38. Date of lumbar pu 39. CSF appearance	performed? ☐ Yes ☐ No ☐ s below. If no, CSF sample sk incture (dd/mm/yyyy) :/☐ Clear and colorless ☐ Cloudnian Colorless ☐ Cloudnian Color	ip to section 3. / 2 0 udy □Blood staine		matic tap 🛚
If yes, complete table 38. Date of lumbar pu 39. CSF	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Corg	ip to section 3. / 2 0 udy □ Blood staine ganism seen □ N	d □Frank blood/trau ot done	matic tap 🛚
If yes, complete table 38. Date of lumbar pu 39. CSF appearance	s below. If no, CSF sample sk uncture (dd/mm/yyyy) :/ \[\text{Clear and colorless} \] \[\text{Clord} \] Unknown	ip to section 3. / 2 0 udy □ Blood staine ganism seen □ N		matic tap
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Corganism seen Corganism seen, describe the 4 hours of the lumbar punctions.	ip to section 3.	ot done	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless	ip to section 3.	ot done	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Corganism seen Corganism seen, describe the 4 hours of the lumbar punctions.	ip to section 3.	ot done	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within Test	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Corganism seen Corganism seen, describe the 4 hours of the lumbar punctions.	ip to section 3.	ot done plood glucose if laborat	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done olood glucose if laborat □other:	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein 42. CSF glucose	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done blood glucose if laborat □other: □other:	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein 42. CSF glucose 43. Plasma glucose a	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done blood glucose if laborat □other: □other:	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein 42. CSF glucose 43. Plasma glucose a time of LP*	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done blood glucose if laborat □other: □other: □other:	
38. Date of lumbar put 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein 42. CSF glucose 43. Plasma glucose atime of LP* 44. CSF RBC count	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done blood glucose if laborat blood glucose if laborat	
If yes, complete table 38. Date of lumbar pu 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein 42. CSF glucose 43. Plasma glucose a time of LP* 44. CSF RBC count 45. CSF WBC count 46. Lymphocytes	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done olood glucose if laborat other: other: other: other: other: other:	<u> </u>
38. Date of lumbar put 39. CSF appearance 40. Gram stain *Must be taken within Test 41. CSF protein 42. CSF glucose 43. Plasma glucose atime of LP* 44. CSF RBC count	s below. If no, CSF sample skincture (dd/mm/yyyy):/ Clear and colorless Clorunknown No organism seen Organism seen the describe the seen at hours of the lumbar punction.	ip to section 3.	ot done blood glucose if laborat blood glucose if laborat	<u> </u>







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4) PATHOGEN TESTING

Record all pathogen testing carried out for differential diagnosis. Record all results available from local, regional or other laboratories. For additional sample type, add to other, or copy in additional rows as needed. For additional follow up sampling, copy table.

Sample type	Pathogen	Date of sampling	Method	Results	Methods/Assays used	Comments
		[dd/mm/yyyy]			useu	
49. Guthrie test (Dried blood spot) Only for under 1 year old		//20	□PCR □Serology □Other:			
Guthrie test (Dried blood spot) Only for under 1 year old		//20	□PCR □Serology □Other:			
50. Blood		_/_/	□PCR □Culture □Serology □Microscopy □Other:			
Blood			□PCR □Culture □Serology □Microscopy □Other:			
51. Urine			□PCR □Culture □Serology □Microscopy □Other:			
Urine			□PCR □Culture □Serology □Microscopy □Other:			
52. □Saliva swab 53. □Throat swab 54. □Nasal			□PCR □Culture □Other:			







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55. CSF		□PCR □Culture □Serology □Microscopy □Other:		
CSF		□PCR □Culture □Serology □Microscopy □Other:		
56. Stool / Feces		□PCR □Culture □Serology □Microscopy □Other:		
Stool/Feces		□PCR □Culture □Serology □Microscopy □Other:		
57. Other (specify):		□PCR □Culture □Serology □Microscopy □Other:		
Other (specify):		□PCR □Culture □Serology □Microscopy □Other:		
Other (specify):		□PCR □Culture □Serology □Microscopy □Other:		
Other (specify):		□PCR □Culture □Serology □Microscopy □Other:		







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5) CASE REPORT	EODM C	OMDI ETED B	v			
	TORIVI C	OWIPLETED B	<u> </u>			
Name and role						
					T	
Signature			Dat	e (dd/mm/yyyy)		